

Original Research Article

Black Sea fish and shellfish as essential source of vitamin B₁₂

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Received: 31 May 2018

Accepted: 28 June 2018

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ABSTRACT

Background: Vitamins are very important for the human body because they are necessary for many biological absorption processes of other nutrients, for cells and tissues growth and repair. Because of this, there are many recommendations on daily vitamins intake, approved by the different country and world food organizations. Usually under the term "vitamin B₁₂" is understood only cyanocobalamin, but actually this name is general and covers all potentially biologically active "cobalamins"- a group of cobalt-containing compounds. Animal foods, as different meat, milk, eggs, fish and shellfish are considered as the main dietary sources of this vitamin. They contain different forms (methylcobalamin and deoxyadenosylcobalamin) of vitamin B₁₂ in different amounts. There is limited information in the scientific literature about the vitamin B₁₂ content in black sea fish and shellfish. The aims of the present work were to determine and compare vitamin B₁₂ contents as well as relative daily intake of vitamins in different fish and shellfish species from Black Sea waters.

Methods: Vitamin B₁₂ was analysed spectrophotometrically. The method for quantitative analysis includes extraction from the edible tissue and enzymatic hydrolysis to release the cobalt ions.

Results: The quantities of vitamin B₁₂ found in the edible tissue of the analysed samples ranged from 0.63 µg.100 g⁻¹ ww to 21.5 µg.100 g⁻¹ ww.

Conclusions: The observed results confirm that all fish and molluscs samples deliver significant quantities of the water-soluble vitamin B₁₂.

Keywords: Cyanocobalamin, UV-Vis, Spectrometry, Cobalt-complexes, Mussel

INTRODUCTION

Vitamin B₁₂ is essential for the human body - the synthesis of red blood cells, DNA, some neurotransmitters and metabolites. It is responsible for maintaining the activity of the immune system, supports a number of processes in the central and peripheral nervous system and plays a key role in the implementation of key processes and protects against many diseases.

All water-soluble vitamins (B-group and C), with the exception of vitamin B₁₂, were found in organisms of plant origin. Sources of biologically active forms of

cobalamins were only organisms of animal origin and some bacteria. Vegetable foods were considered not to contain biologically active forms of vitamin B₁₂.¹ Compared to those of other vitamins, the vitamin B₁₂ molecule is the largest (molecular mass of 1355.4 g.mol⁻¹) and the most complex. The term "vitamin B₁₂" is a generic name that includes all potentially biologically active forms - "cobalamines" (a group of cobalt-containing compounds).² As the main sources of vitamin B₁₂ were considered meat, milk, eggs, and fish and crustaceans.^{3,4} They mainly contain two forms of vitamin methylcobalamine and 59-deoxyadenosylcobalamine, which function as coenzymes in methionine biosynthesis,

as well as in the metabolism of fatty acids in mammalian cells.^{2,5} The absorption and bioavailability of vitamin B₁₂ in the body strongly depends on the gastrointestinal absorption of the different individuals.¹ On consuming or supplementing with excessive doses, the amount that exceeds absorption body's ability was release through the urine.⁶ Most often, vitamin B₁₂ deficiency in the human body was usually caused by malabsorption, combined with inadequate dietary intake.⁷⁻⁹

Studies conducted among a vegan group indicate that they need additional vitamin B₁₂ intake because they do not consume foods of animal origin.⁸ Vegetarians do not suffer from a severe lack because they regularly consume dairy products and eggs, which contains some of the daily vitamin B₁₂ needed.

The need from vitamin B₁₂ quantitation is based on the requirement of clinical analysis (control of blood serum and plasma levels), food analysis (amount in different food matrices), and pharmaceutical substance analysis. Vitamin B₁₂ can be quantitated by various methods such as microbiological, colorimetric, spectrophotometric, atomic absorption, atomic emission and liquid chromatographic.¹⁰ The choice of a suitable method for quantitative analysis depends on different factors: sample type, purpose of analysis, pre-treatment procedure, time of analysis, type and quantities of reagents, sensitivity of the method, and the price. The most commonly used method for quantification of vitamin B₁₂ in complex matrices like blood and serum, characterized by high resolution and sensitivity, is high performance liquid chromatography (HPLC).^{11,12} However, in routine food analysis, the spectrophotometric method also provides the necessary sensitivity. It is characterized by speed, high sensitivity and accuracy, and relatively low cost. The analysis is to determine the content of cobalt ions in the aqueous sample extract by means of a colorimetric reaction.¹³ The quantitative analysis of the vitamin B₁₂, in the selected fish and shellfish species, was performed by a spectrophotometric method.

There is limited information in the scientific literature about the cyanocobalamin's content in Black Sea fish and shellfish. Because of this, the aims of the present work were to determine and compare vitamin B₁₂ contents as well as relative daily intake of vitamins in different fish and shellfish species from Black Sea waters.

METHODS

Reagents and instrumentation

All used reagents were with analytical grade purity, purchased from Sigma Aldrich – cobalt (II) chloride (CoCl₂), acetate buffer (0.1 M, pH 4.0), sodium cyanide (NaCN), α -amylase, pepsin, pyrogallol red, nitric acid (conc. HNO₃), sulfuric acid (conc. H₂SO₄), water and ethanol. For the purposes of the experiment, from the listed reagents were prepared the following solutions: 1

mg.ml⁻¹ cobalt (II) solution in 0.09 M nitric acid, 0.02% pyrogallol red, 1% NaCN.^{12,14}

The spectrophotometric analysis of vitamin B₁₂ was performed on a Hach Lange DR3900 spectrophotometer. It is a single-ray instrument operating in the visible area of electromagnetic spectrum. The quantitative determination of the Co (II) content in the aqueous extracts of the analyzed samples was carried out by measuring their absorption. For this purpose, the spectrophotometer was calibrated by measuring the intensity of Co (II) and pyrogallol red complex in solutions with increasing concentration. Upon construction of the calibration curve, five different standard solutions of CoCl₂ in aqueous nitric acid were used in a concentration range of 0.025-0.4 μ g/ml.

The colored complex of each of the five solutions was synthesized directly in the glass cuvette of the apparatus. Their absorption was measured at $\lambda_{max} = 475$ nm. In the cuvette were mixed 1 ml of the corresponding cobalt solution and 0.5 ml of a 0.02% solution of pyrogallol red. The absorption was read as measured against a blank sample (distilled water with 0.5 ml pyrogallol red solution). The described procedure was used to measure the absorptions of the 5 standard solutions of Co (II). With the obtained data was constructed the calibration curve (Figure 1). The shown linearity of the method was 0.9846.

Samples

All samples - ten Black Sea fish (garfish, sprat, grey mullet, horse mackerel, bonito, bluefish, goby, turbot, shad and red mullet) and shellfish (black mussel and rapana) were used as samples for the present study (table 1). The fish and mollusk species were purchased from Varna local fish markets. All samples were immediately frozen and stored at -20°C in a home fridge. Biometric characteristics as mean weight (g) and mean length (cm) were determined and presented in Table 1.

Samples preparation

Before quantitative spectrometric analysis of vitamin B₁₂, it is necessary to extract from samples. The enzymatic hydrolysis is a traditionally used method for complex matrices. Heudi et al used the enzymes α -amylase and pepsin, 1% NaCN, at pH ~ 4 acetic acid buffer.¹² Under these conditions the samples were hydrolyzed for 3 hours at 37°C. Thereafter, the enzymes were deactivated by prolonged (35 min) heating at 100 - 120° C.

The samples were cooled to room temperature in an ice water bath, filtered through a medium-speed "white blend" filter. The filtrates were used in the subsequent steps of the experiment. The solutions, containing the sample extract of vitamin B₁₂, were subjected to acid decomposition of the cobalamine molecule for the separation of cobalt ion.¹⁵ A mixture of conc. HNO₃:

conc. $H_2SO_4 = 10:1$ was added to a portion of the collected filtrate and the resulting solution was evaporated to near dryness. The residue was neutralized and transferred to a measuring flask and distilled water was added to the mark. The solution was used to perform a spectrophotometric quantitative analysis of the cobalt

ions content. Four parallel samples were processed for each fish species, mussel and rapana (two with and another two without a standard additive). The evaluation of the extraction procedure was carried out using the method of standard addition.

Table 1: Biometrical and biological characteristics of analyzed fish and shellfish.

No	Species	n	Food/habitat	Length, cm mean±SD	Weight, g mean±SD
1.	Sprat (<i>Spratus sprattus sullinig</i>)	21	Zooplankton/pelagic	11.5±0.2	10.5±0.3
2.	Red mullet (<i>Mullus barbatus</i>)	18	Zooplankton and crustaceans/demersal	14.5±1.5	39.3±3.0
3.	Horse mackerel (<i>Trachurus Mediterraneus</i>)	18	Carnivore/pelagic	14.9±1.5	22.9±2.3
4.	Goby (<i>Neogobius melanostomus</i>)	16	Carnivore/demersal	16.7±1.1	69.4±2.4
5.	Shad (<i>Alosa pontica</i>)	7	Carnivore/ pelagic	26.8±2.1	325.0±5.0
6.	Bluefish (<i>Pomatomus saltatrix</i>)	12	Carnivore/pelagic	18.5±1.1	60.0±3.0
7.	Grey mullet (<i>Mugil cephalus</i>)	7	Herbivore/ pelagic	32.0±2.5	290±4.5
8.	Garfish (<i>Belone belone</i>)	7	Carnivore/pelagic	35.0±1.2	52.0±3.5
9.	Bonito (<i>Sarda sarda</i>)	5	Carnivore/pelagic	40.0±1.5	420.0±5.5
10.	Turbot (<i>Pseta maxima</i>)	3	Carnivore/demersal	45.0±2.0	1400.0±10.0
11.	Black mussel (<i>Mytilus galloprovincialis</i>)	50	Herbivore/demersal	5.5±0.5	12.0±0.5
12.	Rapana (<i>Rapana venosa</i>)	15	Carnivore/demersal	10.0±0.5	155.0±5.0

n=number of specimens.

Spectrophotometric analysis

Quantitative determination of cobalt ions in samples was performed analogously to standards. The analyzed solution was prepared directly in the spectrophotometer's cuvette. Combine 1 ml of sample and 0,5 ml of 0,02% pyrogallol red solution. The absorption of resulting complex was measured against a blank sample (distilled water with 0.5 ml pyrogallol red). The amount of cobalt ions was determined by the built-in standard curve.

RESULTS

Table 2: Vitamin B₁₂ content in black sea fish and shellfish.

No	Species	Vitamin B ₁₂ , $\mu\text{g}.100\text{g}^{-1}\text{ww}$
1	Sprat	0.75 ± 0.07
2	Horse mackerel	5.75 ± 0.48
3	Goby	21.50 ± 4.60
4	Grey mullet	5.70 ± 0.80
5	Shad	18.88 ± 3.70
6	Bluefish	13.20 ± 3.10
7	Bonito	6.05 ± 1.40
8	Turbot	19.50 ± 4.30
9	Garfish	0.63 ± 0.08
10	Red mullet	17.40 ± 2.20
11	Black mussel	22.35 ± 3.81
12	Rapana	19.05 ± 2.60

Quantitative determination of the content of Co (II) in ten fish species and Black Sea mussel and rapana was carried

out. The data obtained from the analysis were recalculated and they are shown in Table 2 as micrograms vitamin B₁₂ per 100 grams of raw edible tissue ($\mu\text{g}.100\text{g}^{-1}\text{ww}$) of the test specimens (mean±standard deviation).

DISCUSSION

The shown data variety was in the range of 0.63 $\mu\text{g}.100\text{g}^{-1}\text{ww}$ to 22.35 $\mu\text{g}.100\text{g}^{-1}\text{ww}$. Half of the fish analyzed species showed a content of this vitamin below 10 $\mu\text{g}.100\text{g}^{-1}\text{ww}$, the lowest values were calculated of 0.75 $\mu\text{g}.100\text{g}^{-1}\text{ww}$ in sprat and 0.63 $\mu\text{g}.100\text{g}^{-1}\text{ww}$ in garfish. With exception of shad (18.88 $\mu\text{g}.100\text{g}^{-1}\text{ww}$), all other fish with high levels of vitamin B₁₂ were demersal species - 21.5 $\mu\text{g}.100\text{g}^{-1}\text{ww}$ for goby, 19.50 $\mu\text{g}.100\text{g}^{-1}\text{ww}$ for turbot, 17.40 $\mu\text{g}.100\text{g}^{-1}\text{ww}$ for red mullet and rapana (19.05 $\mu\text{g}.100\text{g}^{-1}\text{ww}$) and mussel (22.35 $\mu\text{g}.100\text{g}^{-1}\text{ww}$).

There are only few studies in the scientific literature presenting results for cyanocobalamin content in raw edible fish or shellfish tissues.^{11,16} Ahmadiania and collective investigated vitamin B₁₂ content in grey mullet fish and the kilka (similar to sprat fish). The found amounts were significantly higher than our data - 13 $\mu\text{g}.100\text{g}^{-1}\text{ww}$ for grey mullet and 5.7 $\mu\text{g}.100\text{g}^{-1}\text{ww}$ for kilka.

Lebiedzinska and colleagues studied the vitamin B₁₂ content in salmon's edible tissue. The presented result (2.8 $\mu\text{g}.100\text{g}^{-1}\text{ww}$) was similar to our data, but falls within the low amount value.¹¹

Some databases also present results for the cyanocobalamin content in fish and shellfish - whole food catalog and USDA food composition database. The first of them shows data for bluefish ($5.39 \mu\text{g}\cdot 100 \text{g}^{-1}$ ww), shad ($0.15 \mu\text{g}\cdot 100 \text{g}^{-1}$ ww) and turbot ($2.20 \mu\text{g}\cdot 100 \text{g}^{-1}$ ww).¹⁷ The results were several times higher than ours.

The USDA food composition database shows quantities of vitamin B₁₂ for eel, mackerel and turbot – $3.00 \mu\text{g}\cdot 100 \text{g}^{-1}$ ww, $13.67 \mu\text{g}\cdot 100 \text{g}^{-1}$ ww and $2.20 \mu\text{g}\cdot 100 \text{g}^{-1}$ ww, respectively.¹⁸ The presented values for eel (like garfish) and mackerel were close to those obtained in our study, while the content of the shad was lower. The data for vitamin B₁₂ content in rapana are very scarce in the scientific literature. The Japanese database presents a result for the amount of vitamin B₁₂ in rapana tissue which is close to that in our research.

Results for cyanocobalamin in mussel were published by Ramasamy Santhanam in the book *Nutritional Marine Life*, as well as in the American database *The Self Nutrition Data*. Their results are close to those found in our study – $12 \mu\text{g}\cdot 100 \text{g}^{-1}$ ww and $17 \mu\text{g}\cdot 100 \text{g}^{-1}$ ww, respectively.¹⁹

The estimation of the edible tissue of the analyzed Black Sea fish and shellfish as a source of vitamin B₁₂ was based on the comparison of the specified amount of vitamins in the recommended daily intake tables for Bulgaria.²⁰ The comparison was made for the age group up to 60 years.

The results shown in Figure 1 represent a percentage of the recommended daily intake (RDI) for vitamin B₁₂ (for 100 grams of edible tissue).

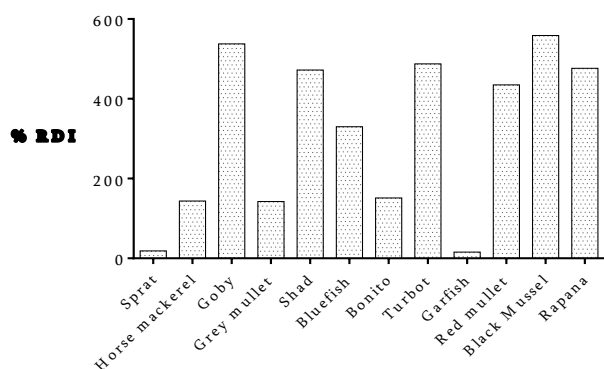


Figure 1: Percentage of the RDI* of vitamin B₁₂ in Black Sea fish and shellfish.

*Relative daily intake (RDI).

Almost all of analyzed fish and shellfish, excluding sprat and garfish, provide significant amounts of vitamin B₁₂ above the RDI. One hundred grams of edible tissue of gray mullet, bonito and horse mackerel contain almost doubled amounts of RDI. In red mullet, turbot, shad,

goby, black mussel and rapana, the quantities were almost seven times higher than the required daily intake.

CONCLUSION

The vitamin B₁₂ content was studied in twelve Black Sea samples of animal origin - ten fish and two molluscs. With the exception of two fish species (sprat and garfish), all others provide significant amounts of the analyte. With a significantly higher content of cyanocobalamin were distinguished all demersal fish, rapana and mussel. Recalculated with RDI, they provide over 500% of the required daily amount of vitamin B₁₂. Bluefish, gray mullet, bonito and horse mackerel contain much less cyanocobalamin, but they also provide significantly more than 100% RDI.

Funding: No funding sources

Conflict of interest: None declared

Ethical approval: Not required

REFERENCES

1. Watanabe F. Vitamin B₁₂ Sources and Bioavailability. *Exp Biol Med.* 2007;232:1266–74.
2. O’Leary F, Samir S. Vitamin B₁₂ in Health and Disease. *Nutrients.* 2010;2:299-316.
3. Scott J. Bioavailability of vitamin B₁₂. *Eur J Clin Nutr.* 1997;51(1):49-53.
4. Watanabe F, Bito T. Vitamin B₁₂ sources and microbial interaction. *Exp Biol Med (Maywood).* 2018;243(2):148-58.
5. Farquharson J, Adams J. The forms of vitamin B₁₂ in foods. *Br J Nutr.* 1976;36(1):127-36.
6. Food and Nutrition Board: Institute of Medicine. Vitamin B₁₂. In *Dietary reference intakes for thiamin, riboflavin, niacin, vitamin B6, folate, vitamin B12, pantothenic acid, biotin and choline.* National Research Council, Ed. Washington, DC, USA: National Academy Press; 1998: 306-356.
7. Ribarova F. *Food and vitamins.* Sofia, Bulgaria 2007.
8. Weir DG, Scott JM. Cobalamins physiology, dietary sources and requirements. In: Sadler M, Strain JJ, Caballero B, eds. *Encyclopedia of human nutrition.* Volume 1. San Diego, CA: Academic Press; 1998;394–401.
9. Hill HAO, Pratt JM, Thorp RG, Ward B, R. Williams JP. The Chemistry of Vitamin B₁₂ - the coordination of biologically important molecules. *Biochem J.* 1970;120:263-9.
10. Karmi O, Zayed A, Baragethi S, Qadi M, Ghanem R. Measurement of vitamin B₁₂ concentration: a review on available methods. *OAB-India.* 2011;2(2):23-32.
11. Lebidzinska A, Marszall ML, Kuta J. Reversed phase high performance liquid chromatography method with coulometric electrochemical and ultraviolet detection for the quantification of

- vitamins B₁, B₆ and B₁₂ in animal and plant foods. *J Chromatogr A.* 2007;1173:71-80.
12. Heudi O, Kilinc T, Fontannaz P, Marley E. Determination of vitamin B in food products and in premixes by reversed-phase high performance liquid chromatography and immunoaffinity extraction. *J Chrom A.* 2006;1101:63-8.
 13. Divarova V, Racheva P, Lekova V, Gavazov K, Dimitrov A. Spectrophotometric determination of cobalt(II) in a liquid-liquid, extraction system containing 4-(2-thiazolylazo) resorcinol and 2,3,5-triphenyl-2h-tetrazolium chloride. *J Chem Tech Met.* 2013;48(6):623-30.
 14. Chohan ZH, Farooq MA. Spectrophotometric Determination of Cobalt(II) Using N-(2-pyrrolylmethylene)- 2,4, 6-triamino-1,3,5- triazine as a Chromogenic Reagent. *J Chem Soc Pak.* 2001;23(3):168-70.
 15. Eskandari H, Ghaziaskar HS, Ensafi AA. A sensitive and simple extractive-Spectrophotometric method for the determination of microgram amount of cobalt by using a-Benzilmonoxime. *Anal Sci.* 2001;17:327-31.
 16. Ahmadnia A, Sahari MA, Barzegar M, Seyfabadi SJ, Abdollahi M. Vitamins Contents of some Commercially Important Fish Species from South Caspian Sea. *Am-Eur J Sust Agr.* 2008;2(3):285-293.
 17. Whole Food Catalog, 2010. Available at: <http://wholefoodcatalog.info/nutrients/>. Accessed on 21 May 2016.
 18. USDA Food Composition Databases, 2018. Available at: <https://ndb.nal.usda.gov/ndb/nutrients/index>. Accessed on 20 May 2018.
 19. Santhanam R. *Nutritional Marine Life.* CRC Press 15.12.2014.
 20. Ordinance no 1 / 22.01.2018 г., Ministry of health, the physiological norms of nutrition of the population. Available at: http://www.mh.government.bg/media/filer_public/2018/02/13/nared_ba1-22-01-2018-fiziologicni-normi-hranene-naselenie.pdf. Accessed on 20 May 2018.

Cite this article as: Dobrev DA, Merdzhanova AV, Stancheva MD, Terziyski DI, Panayotova VZ. Black Sea fish and shellfish as essential source of vitamin B₁₂. *Int J Sci Rep* 2018;4(8):199-203.