Original Research Article

Hepatoprotective effects of partially purified fractions of *Senna occidentalis* ethanolic extract on diethyl nitrosamine-induced toxicity in Wistar rats

Oijochenemi E. Yakubu¹, Eleojo B. Ojogbane², Francis O. Atanu³*, Chukwuka S. M. Udeh⁴, Morayo E. Ale¹, Blessing H. Bello¹

¹Department of Biochemistry, Faculty of Pure and Applied Sciences, Federal University Wukari, Taraba State, Nigeria
²Department of Medical Laboratory Science, Taraba State University Jalingo, Taraba State, Nigeria
³Department of Biochemistry, Faculty of Natural Sciences, Kogi State University, Anyigba, Kogi State, Nigeria
⁴Department of Biochemistry, Faculty of Biological Sciences, University of Nigeria, Nsukka, Nigeria

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*Correspondence:
Dr. Francis O. Atanu,
E-mail: atanufo@gmail.com

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ABSTRACT

**Background:** Induction of toxicity using nitrosamines provides a reliable animal model for the study of oxidative damage to lipids, cellular membranes, proteins and DNA. In the present report, the effects of partially purified fractions of *Senna occidentalis* leaves on diethylnitrosamine intoxicated rats were studied.

**Methods:** Fractions obtained from eluting the column with solvents of increasing polarity, n-hexane, chloroform, ethyl acetate, ethanol, methanol and distilled water were subjected to *in vitro* for their ability to scavenge 1,1-diphenyl-2-picryl hydrazyl (DPPH) radical. Fraction 6a eluted with ethyl acetate:ethanol (50:50) possessed the highest antioxidant activity, this fraction was therefore selected for in vivo studies. Twenty rats, each weighing between 150 to 250 g were randomly allocated into four groups of five rats each. Hepatotoxicity was induced using a single intraperitoneal injection of diethylnitrosamine (DEN) at the 200 mg/kg body weight. Treatment was carried out for 3 weeks by oral gavage as follows: group A, normal control, group B, DEN control, group C, DEN+fraction (10 mg/kg), group D, DEN+silymarin (5 mg/kg).

**Results:** The results showed that DEN toxicity significantly (p<0.05) increased alanine transaminase (ALT) and aspartate transaminase (AST) activities and increased the level of thiobarbituric acid reactive substance (TBARS) in the liver. In contrast, the levels of bilirubin, total protein (TP) and albumin (ALB) decreased. However, treatment of rats with the extract significantly (p<0.05) reduced the concentrations of TBARS, ALT, AST and bilirubin, but increased the concentration of TP and ALB.

**Conclusions:** These results show hepatoprotective potentials of the fraction. Furthermore, GC-MS fingerprinting of fraction 6a revealed the presence of compounds with anticancer, antioxidant and anti-inflammatory properties confirming its high chance for exploration as a medicinal agent.

**Keywords:** Hepatoprotective, *Senna occidentalis*, Diethylnitrosamine, Toxicity, Antioxidant

INTRODUCTION

Natural products of plant origin have been used for the treatment of various diseases for centuries. In developing countries where orthodox medicine is still not affordable to the majority of rural dwellers, herbal medicine is a promising alternative. Consequently, research in phytomedicine (herbal medicine) has led to the discovery of numerous novel therapeutic compounds with prophylactic and curative potentials for several diseases.
including cancer, diabetes, infectious diseases and hepatic injuries.\textsuperscript{1}

The toxic properties of various nitrosamines in animals and humans are well established through numerous adequate experimental investigations. The parenteral or prolonged oral administration of minute quantities of nitrosamines results in severe tissue damage in organs such as the liver arising from intense neutrophilic infiltration, extensive centrlobular hemorrhagic necrosis, bile duct proliferation, fibrosis and bridging necrosis that ends in hepatocarcinogenesis.\textsuperscript{2}

Cancer is a genetic and non-infectious disease resulting in uncontrolled proliferation of cells. In fact advanced stages of cancer are characterised by metastasis of damaged cells to other parts of the body. Cancers affecting the liver result from exposure to environmental toxicants such as DEN account for a substantial source of death around the world.\textsuperscript{3} Though there are orthodox drugs to manage cancer, none can completely treat cancer and are typically accompanied by numerous side effects. Previous several studies have shown that secondary metabolites like polyphenols, terpenes and alkaloids present in plant preparations confer anti-mutagenic and anticancer properties.\textsuperscript{3}

\textit{S. occidentalis} is an herbal plant belonging to the leguminosae family that is commonly found within the tropical and sub-tropical regions of the world, used traditionally for numerous therapeutic reasons.\textsuperscript{4} Experimental reports published in literature suggest that that \textit{S. occidentalis} has antimicrobial, larvicidal, pupicidal, antityransomal activity. Also, there is substantial evidence to show that the plant has antianxiety, antidepressant, antidiabetic and anti-inflammatory properties. The present study investigated the antioxidant and hepatoprotective effects of chromatographic fractions from leaves extract of \textit{S. occidentalis}.

\section*{METHODS}

\subsection*{Sample collection and preparation}

Fresh and healthy looking leaves of \textit{S. occidentalis} leaves were collected from Wapan Nngakwu area of Wukari in Taraba State, Nigeria. The leaves were identified at the herbarium of the department of biological science, federal university Wukari, Taraba State. Leaves were washed with copious amount of water and dried to constant weight. The dried leaves were pounded to fine powder with mortar and pestle and then stored and labeled in dry containers until needed. Only healthy plants were used as the leaves were examined to be free from diseases.

\subsection*{Ethanolic extraction}

The pulverized leaves were soaked in sufficient volume of ethanol for 48 hours at room temperature. It was stirred at 8 hours interval. After 48 hours, the extracts were filtered out first using a clean white sieving mesh and then using Whatman no. 1 filter paper. The filtrate was then concentrated to dryness at 40°C under reduced pressure on a rotary evaporator, extracts were transferred to air-tight containers, corked and preserved in the refrigerator at 4°C until required. Aliquots of the crude plant extract were weighed and used for phytochemical screening and a portion was given to the Wistar rats of the selected groups.

\subsection*{Animal procurement and treatment}

Male and female albino rats (Wistar strain), weighing 150-350 g were obtained from a commercial breeder in Obudu area in Cross Rivers State. The rats were acclimatized for a period of 14 days after randomization, under standard laboratory conditions (25±2°C, relative humidity of 60±5% and 12 hours light/dark cycle). Before and during the experiment, rats were maintained on a commercial feed and drinking water ad libitum.

\subsection*{Fractionation of ethanolic extract}

The ethanol extract was subjected to column chromatography using silica gel stationary phase. The column was eluted using varying solvent combinations of increasing polarity.

\subsection*{Packing of column}

The packing of the column was done according to the method of Yakubu et al.\textsuperscript{5} The lower part of the glass column was soaked with glass wool with the aid of glass rod. Silica gel slurry was prepared by dissolving 235 g of silica gel (mesh size) in 255 ml of absolute n-hexane. The slurry was packed in 30x400 mm chromatographic column. The chromatographic column (30 mm diameter by 400 mm height) was packed with silica gel and the free flow of the solvent was allowed into a conical flask below. The setup was then seen to be in order when the solvent drained freely without carrying either the silica gel or glass wool into the tap. At the end of the packing process, the tap was locked and the column was then allowed for 24 hours to stabilize after which, the clear solvent at the top of the silica gel was allowed to drain down the silica gel meniscus.

\subsection*{Elution}

The method of Yakubu et al was adopted for the elution.\textsuperscript{5} The extract (2 g) was dissolved in 15 ml absolute ethanol and the solution was applied to a chromatographic column (30 mm diameter by 40 mm height). Phytochemicals were eluted from the plant material using solvent combinations of different polarities. They were n-hexane (100:0); n-hexane:chloroform (50:50); chloroform (100:0); chloroform:ethyl acetate (50:50); ethyl acetate (100:0); ethyl acetate:ethanol (50:50); ethanol (100:0);
ethanol:methanol (50:50); methanol (100:0); methanol:distilled water (50:50); distilled water (100:0).

A measured volume (400 ml) of each solvent combination was poured into the column each time using a separator funnel. Eluted fractions were then collected in aliquots of 200 ml using fraction collection tubes.

**Determination of antioxidant activity using DPPH free radical activity**

The antioxidant activity of fractions of ethanol extract was assayed by the DPPH radical scavenging method described by Karadag et al.6

The assay mixture contained 2 ml of 1.0 mM DPPH radical solution prepared in methanol and 1 ml of standard or extract solution of different concentrations (10-500 µg/ml). The solution was rapidly mixed and incubated in dark at 37°C for 20 minutes. The decrease in absorbance of each solution was measured at 517 nm using a spectrophotometer. Ascorbic acid was used as control while 2 ml of 1.0 mM DPPH radical solution with 1 ml ethanol was taken as blank.

**Calculation of free radical scavenging (%)**

\[
\% \text{ free radical scavenging activity} = \left( \frac{A_c - A_s}{A_c} \right) \times 100
\]

where,

\[A_c=\text{absorbance of control},\]
\[A_s=\text{absorbance of sample}.\]

The concentration of sample required to scavenge 50% of DPPH free radical (IC50) was determined from the curve of percentage inhibitions plotted against the respective concentrations.

**Lipid peroxidation assay**

Lipid peroxidation was carried out by the method described by Ortuno et al.7 Briefly, 2 g of liver samples was homogenized with 10 ml of thiobarbituric acid solution reagent and heated at 95°C for 15 min in a boiling water bath. The mixtures were cooled in ice-cold water and centrifuged at 1,500xg for 15 min at 4°C. Absorbance readings were measured at 532 nm (Shimadzu model UV-1601, Japan). The procedure was repeated three times and results were reported as µg malondialdehyde per ml sample (µg MDA/ml).

**Experimental animals**

Albino rats were housed in a controlled environment (12 hr light/dark cycle and temperature of ≃25°C) and fed standard commercial rat feed and water ad libitum in the animal facility of the department of biochemistry, federal university Wukari, Nigeria. Animals were humanely handled according to the guidelines if the national institute of health for the care and use of laboratory animals (NIH publications no. 8023, revised 1978).

**Animal grouping and administration**

Twenty rats, each weighing between 150 to 250 g were randomly selected into four groups of five rats each. Hepatotoxicity was induced using a single intraperitoneal dose of DEN at the 200 mg/kg body weight. Treatment was carried out for 3 weeks orally as follows: group A was normal control, group B was DEN control, group C was DEN+fraction 6a (10 mg/kg), group D was DEN+silymarin (5 mg/kg).

**Blood sample collection and preparation for biochemical assay**

At the end of the experimental period (three weeks), blood samples were collected through cardiac puncture from the rats under light chloroform anesthesia and allowed to clot, then centrifuged for 10 minutes at 3000 rpm. Serum was separated and stored in a clinical refrigerator and was used for the determination of liver function parameters.

**Biochemical assay**

The serum levels of ALT, AST, total bilirubin determined colorimetrically as described by Doumas et al, serum concentration of albumin was determined as described by Bartholomew et al.8-10 Total serum protein level was by the method of Fine.11

**Statistical analysis**

Data collected were subjected to one way analysis of variance (ANOVA) using Dancer's multiple range test (DMRT). Results were expressed in mean±standard deviation and differences at p<0.05 were considered significant.

**RESULTS**

**Total antioxidant capacity (TAC) of fractions of S. occidentalis**

The TAC of the chromatographic fractions was evaluated against DPPH radical. The IC50 of the various fractions is presented in Figure 2. The values of IC50 were significantly lower in fractions 2a (20.5 mg/ml) and 6a (19.5 mg/ml) which implies high antioxidant capacity as compared to the other fractions as shown in Figure 2. The result showed that fractions 6a and 2a require smaller concentrations to inhibit free radical chain reaction as compared to the other fractions.
Inhibition of lipid peroxidation by fractions of S. occidentalis

The anti-lipid peroxidation was assessed by determining the concentration of TBARS in oxidatively stressed tissue homogenates that were pre-treated with the selected fractions. The TBARS values revealed that fraction 6a has the highest anti-lipid peroxidation potential with the lowest concentration of 41.97 µg/ml, followed by fraction 2a (47.07 µg/ml) as represented in Figure 3. Fraction 6a significantly inhibited the generation of TBARS in the tissue homogenates.

Effects of fractions of S. occidentalis on liver function

Liver function was assessed by assaying for the activities of ALT, AST, ALB and concentrations of TB, direct bilirubin (DB) and total protein (TP) and total protein concentrations were shown in Table 1. ALT, AST, TB, and DB values increased significantly (p<0.05) in the animals that were induced with DEN but were not treated (group B) as compared to the normal animals whereas, these indices were significantly (p<0.05) reduced in group B when compared to group A, however, these values were significantly raised compared to the normal animals in groups A and B that were treated with fraction 6a and standard drug (silymarin) respectively.

GC-MS profile of fraction 6a of S. occidentalis

The compounds identified from the fraction eluted with ethyl acetate+ethanol are presented in order of decreasing %area as shown in Table 2 with cyclononene having the highest %area. The GC-MS profile of fraction 6a revealed the presence of cyclononene, alpha-D-glucopyranoside, tetra acetyl-d-xylonic nitrile, 8-nonenoic acid, beta-D-glucopyranose, (aminomethyl) cyclopropane, isobutyl nonyl carbonate, 4-tetradecene, dichloroacetic acid, 9-oxabicyclo [6.1.0] nonane, n-heptyl acrylate, methanamine, N-methoxy, 3-hexyn-1-ol, cyclopropane, 1,1-dimethyl and 4-nonene, 5-nitro.

Table 1: Effect of fraction 6a of ethanol extract of S. occidentalis leaves on biochemical parameters of rats intoxicated with DEN.

<table>
<thead>
<tr>
<th>Groups</th>
<th>ALT (IU/l)</th>
<th>AST (IU/l)</th>
<th>TP (mg/dl)</th>
<th>ALB (mg/dl)</th>
<th>TB (mg/dl)</th>
<th>DB (mg/dl)</th>
</tr>
</thead>
<tbody>
<tr>
<td>A=normal control</td>
<td>31.91±5.46ₐ</td>
<td>49.64±4.63ₐ</td>
<td>72.20±1.90ₐ</td>
<td>34.56±1.09ₐ</td>
<td>2.60±0.26ₐ</td>
<td>1.33±0.15ₐ</td>
</tr>
<tr>
<td>B=DEN control</td>
<td>35.28±4.6ₐ</td>
<td>112.77±45.8₁</td>
<td>62.20±1.90ₐ</td>
<td>31.56±1.09ₐ</td>
<td>3.60±0.26ₐ</td>
<td>2.33±0.15ₐ</td>
</tr>
<tr>
<td>C=DEN+fraction 6a (10 mg/kg)</td>
<td>24.18±10.2ₐ</td>
<td>52.01±26.8ₐ</td>
<td>84.50±1.56ₐ</td>
<td>36.48±1.7ₐ</td>
<td>2.68±0.1ₐ</td>
<td>1.60±0.1ₐ</td>
</tr>
<tr>
<td>D=DEN+silymarin (5 mg/kg)</td>
<td>27.74±3.45ₐ</td>
<td>53.07±34.7ₐ</td>
<td>73.96±3.06ₐ</td>
<td>32.60±1.3ₐ</td>
<td>1.94±0.0ₐ</td>
<td>1.09±0.2₀ₐ</td>
</tr>
</tbody>
</table>

N=5; values are presented in mean±standard deviation; alanine transaminase, ALT (IU/l); aspartate transaminase, AST (IU/l); total protein, TP (mg/dl); albumin, ALB (mg/dl); total bilirubin, TB (mg/dl) and direct bilirubin, DB (mg/dl); values with different superscript down the column are significantly different at p<0.05.

Table 2: GC-MS profile for fraction 6a of ethanolic extract of S. occidentalis leaves.

<table>
<thead>
<tr>
<th>S. no.</th>
<th>RT</th>
<th>Compound</th>
<th>Area (%)</th>
<th>Chemical structure</th>
<th>Biological function</th>
</tr>
</thead>
<tbody>
<tr>
<td>1.</td>
<td>55.952</td>
<td>Cyclononene</td>
<td>23.70</td>
<td>C₂₇H₅₆</td>
<td>Cytotoxicity, antimicrobial</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>124.227</td>
<td></td>
</tr>
<tr>
<td>2.</td>
<td>56.378</td>
<td>Alpha-D-glucopyranoside, methyl</td>
<td>10.39</td>
<td>C₂₁H₂₈O₆</td>
<td>Anticancer</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>194.18</td>
<td></td>
</tr>
<tr>
<td>3.</td>
<td>64.907</td>
<td>8-nonenoic acid</td>
<td>6.88</td>
<td>C₁₉H₂₄O₂</td>
<td>Antifungal</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>156.22</td>
<td></td>
</tr>
<tr>
<td>4.</td>
<td>50.912</td>
<td>Beta-D-glucopyranose</td>
<td>5.51</td>
<td>C₁₉H₂₀O₆</td>
<td>Anti-diabetic, antihypertensive</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>180.16</td>
<td>antihyperlipidemia.</td>
</tr>
</tbody>
</table>

Continued.
<table>
<thead>
<tr>
<th>S. no.</th>
<th>RT</th>
<th>Compound</th>
<th>Area (%)</th>
<th>Chemical structure</th>
<th>Biological function</th>
</tr>
</thead>
<tbody>
<tr>
<td>5.</td>
<td>35.327</td>
<td>(Amino methyl) cyclopropane</td>
<td>4.02</td>
<td>C&lt;sub&gt;6&lt;/sub&gt;H&lt;sub&gt;11&lt;/sub&gt;N</td>
<td>Antidepressant</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>71.12</td>
<td></td>
</tr>
<tr>
<td>6.</td>
<td>63.783</td>
<td>Isobutyl nonyl carbonate</td>
<td>3.54</td>
<td>C&lt;sub&gt;13&lt;/sub&gt;H&lt;sub&gt;26&lt;/sub&gt;O&lt;sub&gt;3&lt;/sub&gt;</td>
<td>This ester is mainly used in herbal compositions for cosmetic purposes.</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>258.40</td>
<td></td>
</tr>
<tr>
<td>7.</td>
<td>63.046</td>
<td>4-tetradecene</td>
<td>3.25</td>
<td>C&lt;sub&gt;14&lt;/sub&gt;H&lt;sub&gt;30&lt;/sub&gt;</td>
<td>Antibacterial</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>196.37</td>
<td></td>
</tr>
<tr>
<td>8.</td>
<td>62.504</td>
<td>Dichloroacetic acid</td>
<td>2.19</td>
<td>C&lt;sub&gt;4&lt;/sub&gt;H&lt;sub&gt;9&lt;/sub&gt;Cl&lt;sub&gt;2&lt;/sub&gt;</td>
<td>Anticancer, topical chemobalation</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>128.94</td>
<td></td>
</tr>
<tr>
<td>9.</td>
<td>61.418</td>
<td>9-oxabicyclo [6.1.0] nonane</td>
<td>2.07</td>
<td>C&lt;sub&gt;10&lt;/sub&gt;H&lt;sub&gt;28&lt;/sub&gt;O&lt;sub&gt;2&lt;/sub&gt;</td>
<td>Antioxidant</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>252.39</td>
<td></td>
</tr>
<tr>
<td>10.</td>
<td>17.300</td>
<td>n-heptyl acrylate</td>
<td>1.37</td>
<td>C&lt;sub&gt;10&lt;/sub&gt;H&lt;sub&gt;18&lt;/sub&gt;O</td>
<td>Anti-inflammatory, antibacterial</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>170.252</td>
<td></td>
</tr>
<tr>
<td>11.</td>
<td>12.570</td>
<td>Methanamine, N-methoxy</td>
<td>1.14</td>
<td>CH&lt;sub&gt;3&lt;/sub&gt;NHOCH&lt;sub&gt;3&lt;/sub&gt;</td>
<td>It is used to illicit production of methamphetamine.</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>61.08</td>
<td></td>
</tr>
<tr>
<td>12.</td>
<td>15.206</td>
<td>3-hexyn-1-ol</td>
<td>1.05</td>
<td>C&lt;sub&gt;10&lt;/sub&gt;H&lt;sub&gt;18&lt;/sub&gt;O</td>
<td>Antiproliferative</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>98.14</td>
<td></td>
</tr>
<tr>
<td>13.</td>
<td>17.067</td>
<td>Cyclopropane, 1,1-dimethyl-</td>
<td>1.04</td>
<td>C&lt;sub&gt;6&lt;/sub&gt;H&lt;sub&gt;10&lt;/sub&gt;</td>
<td>Antitumor</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>70.13</td>
<td></td>
</tr>
<tr>
<td>14.</td>
<td>61.883</td>
<td>4-nonene, 5-nitro</td>
<td>1.02</td>
<td>C&lt;sub&gt;9&lt;/sub&gt;H&lt;sub&gt;17&lt;/sub&gt;N&lt;sub&gt;2&lt;/sub&gt;O&lt;sub&gt;2&lt;/sub&gt;</td>
<td>Antimicrobial</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>171.24</td>
<td></td>
</tr>
</tbody>
</table>

**Figure 1:** Experimental design for the extraction, fractionation and biological evaluation of *S. occidentalis* leaves.
Figure 2: Total antioxidant capacity of solvent fractions of ethanolic extract of *S. occidentalis* leaves; the most active fractions are shaded in black.

Figure 3: Effect of selected solvent fractions of ethanolic extract of *S. occidentalis* on total lipid peroxides in liver; the most active fractions are shaded in black.
DISCUSSION

The total antioxidant capacity was evaluated using the free radical scavenging activity of the various fractions. The DPPH method was a sensitive means to evaluate the antioxidant potential of a given substance or plant extracts in terms of free radical scavenging ability.\(^\text{12}\) DPPH is a stable nitrogen-centered free radical whose color changes from violet to yellow when it is reduced by either the addition of hydrogen or electron, thus, compounds that are capable of reducing DPPH can be termed antioxidants due to their radical scavenging activity.\(^\text{13}\) It was found that all the chromatographic fractions exhibited radical scavenging activity with fraction 6a showing the highest hydrogen donating tendency than the other fractions as evident in its low IC\(_{50}\) value which represented a high inhibitory percentage. The antioxidant properties determined, showed that the solvents were able to extract substances with antioxidant potency. However, water was less efficient in the extraction than other solvents that were employed. This finding was in agreement with the earlier findings of Fatma et al and Vivek et al where antioxidant properties of plant materials were evaluated using DPPH with considerable inhibitory activity against free radicals.\(^\text{14,15}\)

Lipid peroxidation was extensively implicated in primary toxicological effects and was caused by the generation of reactive species from varying sources including organic peroxides, redox cycling and iron-containing substances. Lipid peroxidation is a free radical-induced oxidative chain reaction in which lipid molecules become oxidized to form lipid peroxides and this reaction can be terminated when the substrate is depleted or by when an antioxidant is added, which makes hydrogen available to reduce peroxy radicals.\(^\text{16}\) The TBARS assay has been previously used to determine the extent of lipid peroxidation in biological systems and in food materials. TBA reacts particularly with malondialdehyde (MDA), a secondary product of lipid peroxidation to produce a red chromogen that can be determined using a spectrophotometer.\(^\text{17}\) In this study, all the fractions were capable of reducing the concentration of MDA. But the highest anti-lipid peroxidation activity was observed in fraction 6a. This finding indicated that the fractions of the extracts might prevent reactive radical species from damaging biological and food systems. The result obtained in this study collaborate with the result of Tuyen et al that showed a significant reduction in the concentration of MDA of minced pork upon treatment with the leaves extract of *Phyllanthus acidus*.\(^\text{18}\)

Administration of DEN significantly elevated the values of liver biomarker enzymes. These enzymes were mostly present in low concentrations in the serum and usually in high concentrations in the liver of animals with intact liver architecture. But once the liver architectural integrity was compromised as a result of lipid peroxidation arising from oxidative stress or viral infections such as hepatitis, the enzymes thus escape from the hepatocytes into the bloodstream where they are not known to play any specific role.\(^\text{19}\) The hike in levels of these enzymes could be a result of hepatic necrosis, damage to the bile duct or even fibrosis which preludes hepatic carcinogenesis. However, the administration of the selected fraction 6a was able to normalise the levels of these indices compared to the normal rats. The normalisation corroborates the total antioxidant capacity of the fraction 6a which was further confirmed by its ability to inhibit lipid peroxidation. Similarly, liver damage affected other normal hepatic physiological mechanisms such as the metabolism of bilirubin and protein.\(^\text{20}\) This was evident in the significant increase in the concentration of bilirubin and the decrease in total protein and albumin whose metabolisms are being regulated by the liver. However, the administration of the extraction fraction considerably reversed the conditions to normalcy.

Gas chromatography-mass spectroscopy (GC-MS) is a very compatible and one of the best methods to identify pure compounds present at less than 1 ng in biological specimens for quantification purposes. The unknown organic compounds in a complex biological mixture can be identified by the interpretation and matching of the test sample spectra with the reference spectra.\(^\text{21}\) In this present study, the GC-MS profile of fraction 6 of *S. occidentalis* leaves ethanolic extract, fraction eluted with ethyl acetate:methanol were presented in order of decreasing % area, cyclononene having the highest % area as shown in the above table. Compounds identified from fraction 6 which have antioxidant, anticancer and anti-inflammatory properties are alpha-D-glucopyranoside, methyl, dichloroacetic acid, 9-oxabicyclo [6.1.0] nonane, n-heptyl acrylate, 3-hexyn-1-ol and many others. The GC-MS observation was consistent with the hepatoprotective effect and antioxidant potency of the fraction as demonstrated in this study, most especially that many earlier studies have always implicated the hepatoprotective effect of plant materials to their antioxidant and anti-inflammatory properties.\(^\text{21}\)

CONCLUSION

In conclusion, the findings from this work portray fractions of ethanol extract of *S. occidentalis* leaves as possible antioxidants and anti-hepatotoxic agents that are capable of preventing tissue damage that may be arising from oxidative stress imposed by certain chemicals agents such as DEN. The foregoing properties were collaborated by the presence of some very important bioactive components in the fraction 6a that exhibited a much more promising activity as compared to the other fractions. However, it will be very useful to understand the molecular mechanism of these tissue-protective activities of this fraction.
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Ethical approval: The study was approved by the institutional ethics committee

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